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Tetrahedron 60 (2004) 549–559

Tetrahedron

Dioxygenase-catalysed sulfoxidation of bicyclic alkylaryl sulfides and chemoenzymatic synthesis of acyclic disulfoxides

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Received 8 August 2003; revised 28 August 2003; accepted 17 October 2003

Abstract—Toluene- and naphthalene-dioxygenase-catalysed oxidation of six bicyclic disulfide substrates, using whole cells of Pseudomonas putida, gave the corresponding monosulfoxides with high ee values and enantiocomplementarity, in most cases. Two alcohol-sulfoxide diastereoisomers, formed from the reaction of the (R) -1,3-benzodithiole-1-oxide metabolite with *n*-butyllithium and benzaldehyde, were separated and stereochemically assigned. Treatment, of enantiopure (1R,3R)-benzo-1,3-dithiole-1,3-dioxide, obtained by chemoenzymatic synthesis, with alkyllithium reagents, resulted in a novel ring-opening reaction which proceeded with inversion of configuration to yield a series of acyclic disulfoxides.

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1. Introduction

The potential of microbial enzymes in the biocatalytic oxidation of sulfides, to yield enantiopure sulfoxides, has been recognised for many years. t-Butyl p-tolyl sulfoxide, the first reported enantiopure example of a sulfoxide produced by enzyme-catalysed oxidation of an achiral sulphide, $\frac{1}{1}$ $\frac{1}{1}$ $\frac{1}{1}$ was obtained using the fungus Aspergillus niger (foetidus), a source of monooxygenase enzymes. More recent biotransformation studies, from these $2²⁻⁷$ and other laboratories, $8,9$ have shown that the ring hydroxylating dioxygenase enzymes (Rieske non-heme iron oxygenases), present in the soil bacterium Pseudomonas putida, are among the most stereoselective biocatalysts, currently available, for the production of enantiopure acyclic sulfoxides. Wild-type and mutant P. putida strains, $2-\frac{8}{3}$ and E. coli recombinant strains, $3,5,8,9$ containing toluene dioxygenase (TDO) and naphthalene-dioxygenase (NDO), have thus, to date, yielded >30 sulfoxides with high ($>90\%$) enantiomeric excess (ee) values. $2-9$ Routes to enantiopure sulfoxides are required for asymmetric synthesis studies (e.g. as chiral auxiliaries and ligands) and for the production of pharmaceuticals including anti-ulcer drugs containing a chiral sulfoxide group (e.g. the proton pump inhibitors omeprazole, and lanzaprazole). The results, presented in this

article, provide a further demonstration of the value of enantiocomplementary dioxygenase enzymes for the production of enantiopure cyclic monosulfoxides from 1,3-, 1,4- and 1,5-disulfides. The enantiopure monosulfoxide metabolite 1B, from benzo-1,3-dithiole 1A, and the derived disulfoxide 1C, have, in turn, been used to investigate their reactions with alkyllithium reagents.

2. Results and discussion

2.1. Dioxygenase-catalysed monosulfoxidation of sulfides 1A–8A to yield the corresponding sulfoxides 1B–8B

The initial phase of the study involved biotransformations of bicyclic disulfide substrates 1A–6A using: (i) P. putida UV4 (a constitutive mutant strain containing TDO), (ii) P. putida NCIMB 8859 (a wild-type strain containing NDO) and, to a lesser extent, (iii), P. putida 9816/11 (an inducible mutant strain containing NDO). These whole-cell systems provide sources of the required dioxygenase biocatalysts, under biotransformation conditions similar to those reported for earlier sulfoxidation studies.²⁻⁹ The isolated yields, enantiopurity values, and absolute configurations of monosulfoxides $1B-6B$ are shown in [Table 1](#page-1-0). Sulfoxides $1B$, 10 $2B_{cis}$, ^{[10](#page-10-0)} $2B_{trans}$, ¹⁰ $3B_{cis}$, ¹⁰ $3B_{trans}$ ¹⁰ and $6B$ ^{[11](#page-10-0)} had earlier been reported as single enantiomers, while sulfoxides 4B and 5B were obtained for the first time as single enantiomers, during the present study.

Keywords: Dioxygenase-catalysed sulfoxidation; Enantiopure bicyclic sulfoxides; Alkylithium; Acyclic disulfoxides.

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Table 1. Isolated yields (%Yld.), ee values (% ee) and absolute configurations (Ab.Con.) of sulfoxides 1B–8B obtained from the corresponding sulfides 1A–8A using TDO (P. putida UV4) and NDO (P. putida 8859) in whole-cell systems

Compound	TDO enzyme			NDO enzyme			
	%Yld.	$%$ ee	Ab.Con.	%Yld.	$%$ ee	Ab.Con.	
1В	20	33	S	74, 72	>98	R	
$2B_{cis}$	40, 26	>98	1S, 2R	49	82	1R.2S	
$2B_{trans}$	5	>98	1S,2S	7	38	1R, 2R	
$3B_{cis}$	26	>98	1S, 2R				
$3B_{trans}$	5	18	1R,2R				
4B	1, 3	>98	R	78	>98	$S^{\rm a}$	
5B	$\mathbf b$	$\mathbf b$	$\mathbf b$	26	$80(93)^{c}$	S	
6 B	23	> 98	R	60	> 98	S	
7B	5	26	R	21	38	S	
8B	10	> 98	R	\overline{c}	43	R	

^a Predicted absolute configuration. b No sulfoxide isolated; using *P. putida* 9816/11 as a source of NDO

biocatalyst.
Recrystallisation of bioproduct from P. putida 9816/11 gave an ee value of $>98%$.

The circular dichroism (CD) spectra, of sulfoxide metabolites 1B, $2B_{cis}$, $2B_{trans}$, $3B_{cis}$, and $3B_{trans}$, were found to have a strong absorption around 220 nm due to an n– π^* transition associated with the sulfoxide group.^{[10](#page-10-0)} This trend, of a positive CD absorption in the 220 nm region, was consistently associated with the rigorously established (R) configuration in sulfoxides 1B, $2B_{cis}$, $2B_{trans}$, $3B_{cis}$, and $3B_{trans}$. The absolute configurations of sulfoxide enantiomers 4B–6B have not been reported in the literature; no consistent CD spectral trend was evident that would allow reliable assignment of their absolute configurations. Hence, alternative approaches, to the configuration assignments for sulfoxide metabolites 4B–6B were examined. Earlier studies, of the biotransformation of alkyl- and alkenylphenyl sulfides (RSPh, $R=Me$, Et, $CH₂=CH₂$, etc.), using P. putida UV4 (TDO), showed that an (R) configuration was observed in all cases.^{[5](#page-10-0)} This empirical method, of configuration assignment, was thus based on the assumption that the cyclic alkylaryl and alkenylaryl sulfides 4A–6A would bind in a similar manner, at the active site of TDO, and would also show a preference for $(1R)$ configurations for sulfoxide metabolites 4B–6B (Scheme 1).

Scheme 1.

Conversely, a strong preference for the opposite (S) sulfoxide configuration was observed earlier, using NDO with some alkylphenyl and alkenylphenyl sulfides (RSPh, R=Me, Et, $CH_2=CH_2$, etc.).^{[5](#page-10-0)} X-ray crystallographic evidence, of the substrate binding and catalytic sites of NDO, that can account for the absolute stereochemistry of the cis-diol metabolites of bicyclic alkenes (e.g. indene) and arenes (e.g. naphthalene), has recently been reported.^{[12](#page-10-0)} In view of the similarity in shape between naphthalene and the bicyclic sulfide substrates 4A and 6A it is probable that substrate binding and catalysis, using NDO [\(Scheme 2\)](#page-2-0), will yield bioproducts of similar configuration (but opposite to that obtained using TDO). Thus, absolute configurations of sulfoxide metabolites 4B–6B were tentatively assigned by comparison with earlier results, using the TDO and NDO enzyme systems and acyclic alkylphenyl sulfides as substrates.

Recrystallization of the bioproduct, $(-)$ -1,2,3,4-tetrahydro- $1\lambda^4$,5-benzodithiepin-1-oxide 5B (80–93% ee) obtained by NDO-catalysed sulfoxidation, provided an enantiopure

sample whose absolute configuration was unequivocally established, as (1S) by X-ray crystallography, using the anomalous dispersion method (Fig. 1). The tetrahydrodithiepine ring adopts the chair conformation, with an equatorial oxygen atom. This result lends additional support to the tentative assignments of $(1R)$ and $(1S)$ configurations for alkylaryl sulfoxides 4B–6B formed with TDO and NDO biocatalysts, respectively.

Figure 1. X-ray structure of 5B.

As an extension to the dioxygenase-catalysed sulfoxidation study, the monosulfide isosteres 7A and 8A, of the parent disulfide substrate 1A, were also examined. Thus, the corresponding enantioenriched sulfoxides, 7B and 8B, of known absolute configuration, were obtained with TDO and NDO as biocatalysts [\(Table 1\)](#page-1-0). While sulfoxides 7B, obtained using TDO and NDO, were of opposite absolute configuration, surprisingly, identical $(1R)$ configurations were found for the sulfoxide metabolites 8B using either enzyme. Sulfoxide 7B was the most polar of the five chiral metabolites isolated with P . putida UV4 and 2,3-dihydrobenzo[b]thiophene 7A as substrate. The structure and stereochemistry (absolute configurations and ee values) of the other metabolites, resulting from benzylic hydroxylation, desaturation and dihydroxylation of the heterocyclic and carbocyclic rings, will be discussed elsewhere.

The results, shown in [Table 1,](#page-1-0) indicate that the cyclic sulfoxides were isolated in variable yields $(1-78%)$ and that all but the sulfoxides $3B_{trans}$ and 7B could be obtained with high ee values $(>90\%)$, using either TDO or NDO as biocatalyst. The contrasting stereopreferences, previously

observed for TDO- and NDO-catalysed oxidation of some acyclic alkylaryl sulfides⁵ ([Scheme 1](#page-1-0)), were even more common during the sulfoxidation of cyclic alkylaryl sulfides to yield sulfoxide metabolites, $1B$, $2B_{cis}$, $2B_{trans}$, $4B$, $6B$ and 7B. The most striking examples of enantiocomplementarity were found when TDO or NDO biocatalysts were used to produce enantiopure sulfoxides 4B and 6B of opposite configuration.

The Kagan modified Sharpless reagent, $(Ti (i-Pro)_A/DET/$ t -BuOOH), had earlier¹¹ been used to obtain sulfoxide $6B$, as a single enantiomer (8% yield), after separation from a product mixture containing three other sulfoxidation products (92% yield). Biocatalysis now provides a complementary synthesis (asymmetric oxidation) of sulfoxide 6B where either enantiomer is available in higher yield and state of purity. While the stereoselectivity values, found during sulfoxidations using TDO or NDO, were similar, the isolated yields were generally better, using the NDO system.

Results from earlier biotransformation studies, using both acyclic sulfides, and the corresponding racemic sulfoxides as substrates, for purified dioxygenases (TDO and NDO), or whole cell bacterial strains, containing TDO and $NDO_{5,8}$ $NDO_{5,8}$ $NDO_{5,8}$ suggested that the enantiopure sulfoxide products were exclusively (or mainly) formed by asymmetric sulfoxidation. During the present study, the cyclic racemic sulfoxides 1B–8B were also tested as substrates for the UV4 and 8859 strains of P. putida. With the exception of recovered sulfoxide $8B(1S, 74\%$ ee, 8859 strain), the other biotransformations showed only poor enantioenrichment of residual sulfoxides. On this basis, it would again appear that the high ee values, observed for the sulfoxide metabolites of cyclic sulfides, using TDO and NDO, were mainly the products of asymmetric sulfoxidation rather than kinetic resolution. However, during the course of these biotransformation studies of racemic sulfoxides with P. putida UV4, tentative evidence was found of other reactions occurring $\leq 5\%$ yield of bioproducts). This included evidence of a reductase-catalysed deoxygenation of sulfoxides which will be presented elsewhere.

2.2. Reaction of (R)-benzo-1,3-dithiole-1-oxide 1B with n-butyllithium and benzaldehyde

Small quantities, of enantiopure $(+)$ - and $(-)$ -sulfoxide 1B, had earlier been obtained by chromatographic resolution of the racemate, using semi-preparative chiral stationary phase HPLC (CSPHPLC).^{[10](#page-10-0)} Treatment, of the sulfoxide enantiomers 1B with strong base, potassium bis(trimethysilyl) acetamide, yielded a carbanion at C-2 that was used in the nucleophilic substitution of alkyl halides, to provide twelve enantiopure 2-substituted benzo-1,3-dithiole sulfoxides, including compounds $2B_{cis}$, $2B_{trans}$, $3B_{cis}$, $3B_{trans}$.^{[10](#page-10-0)} Absolute configuration determination, of the derived $(1S, 2S, 2'S)$ -2- $(2⁷$ -methylbutyl)-1,3-benzodithiole 1-oxide, by X-ray crystallography, allowed the configurations of the other 2-substituted benzo-1,3-dithiole sulfoxides 1B, $2B_{cis}$, $2B_{trans}$, $3B_{cis}$, $3B_{trans}$ to be unequivocally assigned by stereochemical correlation and CD spectral comparison.

A major disadvantage of the CSPHPLC resolution procedure was that only small quantities, of each sulfoxide

Scheme 3.

enantiomer 1B (ca: 40 mg), were obtained for reactivity studies.[10](#page-10-0) With the availability of larger samples of the enantiopure benzo-1,3-dithiole sulfoxide 1B, from the NDO-catalysed biotransformation route, it was possible to re-examine the reactivity of this compound and its derivatives. Thus, the second phase of the study involved the reactions of enantiopure monosulfoxide 1B and disulfoxide $1C_{trans}$ with alkyllithium reagents.

(R)-Monosulfoxide metabolite 1B ($[\alpha]_D = +505$) was treated with 1 equiv. of *n*-butyllithium in dry THF $(-78 \degree C)$ followed by the addition of benzaldehyde to the C-2 carbanion intermediate, to yield a diastereoisomeric mixture of 2-substituted benzo-1,3-dithiole sulfoxides 9/10 (Scheme 3). Separation, by PLC, gave cis isomer 10 as a crystalline solid (R_f 0.31, $[\alpha]_D$ =+344, 31% yield) and trans isomer 9 as a viscous oil (R_f 0.19, $[\alpha]_D$ =+116, 23% yield). X-ray crystallographic analysis (Fig. 2) confirmed the cis geometry of sulfoxide 10 and, using the anomalous dispersion method, allowed assignment of absolute configuration, $(1R, 2S, 1'S)$, of the three chiral centres. The heterocyclic ring adopts an envelope conformation, with pseudoaxial oxygen and pseudoequatorial side chain at C(2).

Figure 2. X-ray structure of 10.

The X-ray crystallographic analysis, of the minor diastereoisomer 9, was precluded, since it could not be obtained in crystalline form. Its absolute configuration was, however, assigned by a mild rhenium-catalysed deoxygenation process.^{[13](#page-10-0)} Thus, using the *trans* monosulfoxide 9 and a catalytic amount of $ReOCl₃(PPh₃)₂$ in the presence of triphenyl phosphine (ambient temperature, 1.5 h), the $(-)$ -enantiomer of 2-substituted benzo-1,3dithiole 11 ($[\alpha]_D = -12$, 49% yield) was obtained. Similar treatment (72 h) , of the more hindered *cis* sulfoxide 10, yielded the $(+)$ -enantiomer of 2-substituted benzo-1,3dithiole 11 (α]_D=+12, 44% yield). Since the absolute configuration, of the exocyclic chiral centre of sulfoxide 10, was established as $(1'S)$ by X-ray crystallography, sulfoxide **9** must have the $(1R, 2R, 1/R)$ configuration. The formation, of either the $(1/R)$ or $(1'S)$ enantiomer of alcohol 11, provides a good example, of the transfer of chirality from a $(1R)$ cyclic sulfoxide stereogenic centre in metabolite **1B**, to a new exocyclic chiral centre in alcohol 11. The converse process, i.e. chiral relay from the chiral alcohol centres in an arene cis-dihydrodiol metabolite, to a new chiral sulfoxide centre of either configuration (catechol sulfoxides), has recently been reported from these laboratories.^{[7](#page-10-0)}

The marked degree of diastereoselectivity, found during substitution at the C-2 position for the (R) enantiomer of monosulfoxide 1B, was noteworthy; thus, only two diastereoisomers, $(1R, 2S, 1'S)$ -10 and $(1R, 2R, 1'R)$ -9, of the possible four were detected. Examination of molecular models suggests that this could be due to the preferential attractive interactions between the sulfoxide oxygen atom and the lithium alkoxide, favouring chelate formations $(1R, 2R, 1'R)$ -9^{*} and $(1R, 2S, 1'S)$ -10^{*} (Scheme 4). Similar types of chelate interactions have been suggested, to account for the diastereoselectivity associated with carbanion reactions of trans-1,3-dithiane 1,3-dioxide with benzaldehyde.^{[14](#page-10-0)}

Scheme 4.

2.3. Ring-opening reactions of $(1R,3R)$ -disulfoxide $1C_{trans}$ and monosulfoxides 7B and 8B

Chemical oxidation of $(+)$ - (R) -benzo-1,3-dithiole 1-oxide

1B ($[\alpha]_D$ =+505), using NaIO₄ as oxidant, yielded enantiopure (+)-(1R,3R)-disulfoxide $1C_{trans}$ ([α]_D=+646, 75% yield, [Scheme 3](#page-3-0)) as a major product. Compound $1C_{trans}$ was readily separated from achiral (1R,3S) isomer $1C_{cis}$ (19%) yield) by flash chromatography. It was anticipated that treatment of $(+)$ -disulfoxide $1C_{trans}$ in a manner similar to (+) sulfoxide 1B, i.e. reaction with *n*-butyllithum in dry THF $(-78 \degree C)$, followed by addition of benzaldehyde, would give the two diastereoisomers of alcohol sulfoxide 12 ([Scheme 3\)](#page-3-0). In an earlier study, 14 on the synthesis and reaction of (\pm) -disulfoxide $1C_{trans}$ with *n*-butyllithum and pivalaldehyde, alcohol disulfoxide diastereoisomers of similar structure to compound 12 (Ph replaced by t -Bu) were reported. However, spectral analysis and PLC purification indicated that only one product was formed from the reaction of *n*-butyllithum/benzaldehyde $(-78 \degree C)$ with the (1R,3R) enantiomer of cyclic disulfoxide $1C_{trans}$; it was identified as the acyclic disulfoxide 13 (α _D $=-180$, 49% yield, Scheme 5). When this reaction was repeated under identical conditions, but without addition of benzaldehyde, the acyclic $(-)$ -disulfoxide 13 was again formed (Scheme 5).

Since only a few methods are available for the synthesis of enantiopure acyclic disulfoxides, using either chemical or enzymatic approaches, $15 - 18$ the unusual ring-opening reaction of $(1R,3R)$ -disulfoxide $1C_{trans}$ was examined further (Scheme 5). Reactions of $1C_{trans}$ with other alkyllithium reagents (RLi, R=Ph, Me, $(CH₂)₅Me$, t-Bu) were conducted under identical conditions. ¹H NMR analysis, of the resulting acyclic disulfoxide products 13–17 indicated that in all cases single diastereoisomers were formed, albeit in relatively modest yields (28–65%, Table 2). Compound 14 was the only disulfoxide obtained without either a measurable optical rotation or CD absorption. This observation was consistent with total inversion of configuration, occurring during the ring opening process, to yield achiral meso isomer 14. Comparison of the melting points and ¹H NMR data for disulfoxide 14 (mp $124-125$ °C, $[\alpha]_D=0$, isolated during the present study, with that reported for the *meso* (mp 123 °C, $[\alpha]_D = 0$) and racemic

Table 2. Reactions of bicyclic sulfoxides 1C_{trans}, 7B and 8B with alkyllithium reagents to yield the acyclic disulfoxides 13–17 and monosulfoxides 18 and 19

Reactant sulfoxide		Alkyl lithium		Di- and mono-sulfoxide products		
Structure	Ab.Con.		Number Yield		$\left[\alpha\right]_{D}^{a}$	Ab.Con.
$1C_{trans}$	1R, 3R	BuLi.	13	49	-180	1R,3S
$1C_{trans}$	1R, 3R	PhLi	14	32	h	h
$1C_{trans}$	1R, 3R	MeLi	15	59	-234	1R,3S
$1C_{trans}$	1R, 3R	HexylLi	16	65	-247	1R.3S
$1C_{trans}$	1R, 3R	t -BuLi	17	30	$+75$	1R.3S
7B	R	BuLi	18	28	-13°	R
8B	S	BuLi	19	34	$+74^d$	R

^a CHCl₃ solvent.
^b Achiral compound.
^c Formed from **7B** of 90% ee.
^d Formed from **8B** of 73% ee.

(R/S or S/R) (mp 136 °C) isomeric forms,^{[15,16](#page-10-0)} confirmed that it was, indeed, the meso compound.

The characteristic ¹H NMR spectral data, reported for the racemic $(1R,3R/1S,3S)$ and $(1R,3S/1S,3R)$ diastereoisomeric pairs of sulfoxide [15](#page-10-0),¹⁵ suggested that the ring-opening product 15 ($[\alpha]_D = -234$, 59% yield) was either (1R,3S) or (1S,3R) diastereoisomer. Unfortunately an $\lceil \alpha \rceil_D$ value for either enantiomer of compound 15 has not been reported. The $(1R,3S)$ configuration for $(-)$ -sulfoxide 15 was confirmed by partial deoxygenation, using the rhenium complex-catalysed procedure $(Scheme 6)$.^{[13](#page-10-0)} As expected from earlier studies, 13 the dialkyl sulfoxide group was deoxygenated more slowly compared with the alkylaryl sulfoxide group, leading to a 2:1 ratio of dialkyl sulfoxide 20 and alkylaryl sulfoxide 21. Partial racemisation, of monosulfoxides 20 $([\alpha]_D = -27$, 24% ee) and 21 $(|\alpha|_D = -110, 87\%$ ee), occurring to different degrees during the deoxygenation process, was also observed. Despite the partial racemisation, it was possible to assign (S) and (R) configurations, respectively, to monosulfoxides $(-)$ -20 and $(-)$ -21, by comparison of the $[\alpha]_D$ values with authentic samples; $\frac{18}{18}$ $\frac{18}{18}$ $\frac{18}{18}$ thus, a (1R,3S) configuration was assigned to disulfoxide 15. The CD spectra of single diastereoisomers 13, 15–17, obtained by the ring-opening reaction of disulfoxide $1C_{trans}$ (Scheme 5), displayed a similar pattern, i.e. a strong positive absorption at 218– 220 nm and a strong negative absorption at 246–256 nm. The similarity in CD spectra of compounds 13, 15–17 and unequivocal assignment of absolute configuration to disulfoxide 15 led us to the conclusion that in the ringopening reactions inversion of configuration had occurred. This premise was supported by two reports in the literature,^{[19,20](#page-10-0)} where enantiopure acyclic alkylaryl monosulfoxides were found to react with alkyllithium reagents, to yield dialkyl sulfoxides via an S_N2 exchange process occurring with inversion of configuration. As found for acyclic alkylaryl sulfoxides,^{[19,20](#page-10-0)} the aryl–SO bond in the

Scheme 6.

cyclic disulfoxide $1C_{trans}$ was cleaved in preference to the alkyl–SO bond.

It was reported earlier^{[20](#page-10-0)} that alternative reactions of acyclic monosulfoxides with alkyl lithium reagents result in (a) abstraction of an α -hydrogen atom to give an α -lithiosulfoxide (carbanion formation), and (b) replacement of an aryl sulfoxide group with the alkyllithium group (group exchange). The relative proportion of either reaction was found to depend on the structure of sulfoxide (number and acidity of α -hydrogen atoms, substituent size and leaving group ability) and the type of alkyllithium reagent (size and basicity). Our observation of ring-opening reactions (group exchange), of disulfoxide $1C_{trans}$ with alkyllithium reagents $(-78 \degree C)$, suggests that the temperature may also be an important factor. The reported^{[14](#page-10-0)} substitution reaction of a racemic sample of compound $1C_{trans}$, using *n*-butyllithium and pivalaldehyde to form t-butyl analogues of compounds $(1R, 3R, 1'S)$ -12 and $(1R, 3R, 1'R)$ -12 ([Scheme 3](#page-3-0)), was carried out at higher temperatures (0 to -45 °C). Furthermore, ¹H NMR spectral analysis of the crude product mixture, obtained from the reaction of compound $1C_{trans}$ with n-butyllithium and benzaldehyde at a relatively higher temperature $(-45 \degree C)$ indicated the formation of compounds $(1R, 3R, 1'S)$ -12 and $(1R, 3R, 1'R)$ -12.

When the enantiopure monosulfoxide metabolites $4B-6B$ containing six and seven-membered rings, were converted to the corresponding disulfoxides, and subsequently treated with alkyllithium reagents, no evidence of comparable ringopening reactions was observed. However, under the standard reaction conditions monosulfoxide metabolites 7B and 8B, containing a five-membered ring, were also found to undergo a similar ring opening reaction to give the corresponding acyclic dialkyl monosulfoxides 18 and 19 in low yields (28 and 34%, [Scheme 5,](#page-4-0) [Table 2\)](#page-4-0). Since only limited amounts of enantioenriched/enantiopure sulfoxides 7B and 8B were available from the small-scale biotransformations, additional samples of enantioenriched sulfoxides $(-)$ - (R) -7B (90% ee) and $(-)$ - (S) -8B (73% ee) were synthesised by chemical asymmetric sulfoxidation using the Kagan modified Sharpless reagent $(Ti(i-Pro)_{4}/DET/$ t -BuOOH). 11 11 11

It was assumed that inversion of configuration had occurred, again, during the ring-opening reaction of compound $(-)$ -(R)-7B to yield monosulfoxide 18 (α]_D=-13, 28% yield) and of compound $(-)$ - (S) -8B to give the hemiacetal derivative 19 ($[\alpha]_D$ =+74, 34% yield). Chiral stationary phase HPLC (CSPHPLC) analysis of the sulfoxide 19 (49% ee), obtained from sulfoxide 8B (73% ee), showed that partial racemization had occurred. The ee value for sulfoxide 18 could not be obtained using the latter CSPHPLC method.

3. Conclusions

The dioxygenase enzyme system has been shown to catalyse the enantioselective heteroatom oxidation of bicyclic sulfides $(7A, 8A)$ and disulfides $(1A-6A)$. In many cases, the corresponding enantiopure monosulfoxides (e.g. 1B, $2B_{cis}$, $2B_{trans}$, $3B_{cis}$, $4B-6B$) were obtained with opposite

absolute configurations, using either TDO or NDO as biocatalyst. The monosulfoxide 1B was found to undergo an a-hydrogen atom abstraction process with butyllithium followed by reaction, at low temperatures, with benzaldehyde in a diastereoselective manner, to form alcohols 9 and 10. Under these conditions, disulfoxide $1C_{trans}$ was found to undergo a ring-opening reaction to yield the acyclic disulfoxides. A new chemoenzymatic route to enantiopure/ enantioenriched disulfoxides (13, 15–17) and monosulfoxides (18 and 19) was revealed, by the reactions of sulfoxides 1C, 7B and 8B, with alkyllithium reagents.

4. Experimental

4.1. General

¹H NMR spectra were recorded at 300 MHz (Bruker Avance DPX-300) and 500 MHz (Bruker Avance DRX- 500) in CDCl₃ solvent, unless stated otherwise. Chemical shifts (δ) are reported in ppm relative to SiMe₄; coupling constants (J) are given in Hz. Mass spectra were recorded at 70 eV on a VG Autospec Mass Spectrometer, using a heated inlet system. Accurate molecular weights were determined by the peak matching method with perfluorokerosene as standard. Elemental microanalyses were obtained on a Perkin–Elmer 2400 CHN microanalyser. HPLC analyses were carried out using a Perkin–Elmer Series 3B liquid chromatograph coupled to a Perkin–Elmer LC1-100 computing integrator. Analytical TLC was performed on Merck Kieselgel 60_{254} plastic sheets and preparative TLC (PLC) on glass plates $(20 \text{ cm} \times 20 \text{ cm})$ coated with Merck Kieselgel PF₂₅₄₊₃₆₆. Optical rotation ($[\alpha]_D$) measurements were carried out with a Perkin–Elmer 214 polarimeter at ambient temperature (ca. 20° C) at a concentration of \sim 0.01 g cm⁻³ and are given in units of 10⁻¹ deg cm² g⁻¹. Circular dichroism spectra were recorded on a JASCO J-720 instrument, using spectroscopic grade acetonitrile as solvent. Enantiopurity of sulfoxides was determined by CSPHPLC analysis, using Chiralcel OB, OD and OJ columns and hexane/2-propanol (9/1) as eluent. The NCIMB 8859 strain of P. putida was acquired from the National Cultures of Industrial and marine Bacteria, Aberdeen. The 9816/11 strain of P. putida originated from the laboratories of Professor D. T. Gibson (University of Iowa) and the UV4 strain of P. putida was originally provided by Zeneca (now Avecia Lifescience Molecules, Billingham).

4.2. Synthesis and characterization of compounds 1A–8A

The di- and mono-sulfide substrates $1A-3A$, 10 10 10 $4A$, 21 21 21 $5A$, 22 22 22 $6A²³ 7A²⁴ 8A²⁵$ were synthesised using the reported methods.

4.2.1. 1,3-Benzodithiole 1A. Colourless oil; bp 113– 114 °C/3.5 mmHg; lit.^{[10](#page-10-0)} 105 °C/3 mmHg; $\delta_{\rm H}$ (500 MHz, $CDCl₃$) 4.48 (2H, s, CH₂), 7.00–7.02 (2H, dd, J=5.7, 3.3 Hz, $2 \times Ar-H$), $7.19 - 7.22$ (2H, dd, $J=5.7$, 3.3 Hz, $2\times$ Ar-H).

4.2.2. 2-Methyl-1,3-benzodithiole 2A. Colourless oil; bp 98 °C/0.05 mmHg; lit.^{[10](#page-10-0)} 62–64 °C/0.02 mmHg; $\delta_{\rm H}$

 $(500 \text{ MHz}, \text{CDCl}_3)$ 1.70 (3H, d, $J_{\text{Me,H}}$ =6.8 Hz, Me), 4.98 (1H, q, $J_{H Me}$ =6.8 Hz, CHMe), 7.01–7.03 (2H, dd, J=5.8, 3.2 Hz, $2 \times Ar-H$), $7.21 - 7.23$ (2H, dd, $J=5.8$, 3.2 Hz, $2\times$ Ar-H).

4.2.3. 2-Ethyl-1,3-benzodithiole 3A. Colourless oil; bp 70 °C/0.02 mmHg; lit.¹⁰ 72–76 °C/0.01 mmHg; $\delta_{\rm H}$ $(300 \text{ MHz}, \text{CDCl}_3)$ 1.02 (3H, t, $J_{\text{Me}, \text{CH}_2} = 7.0 \text{ Hz}, \text{ CH}_2 \text{Me}$), 1.90–1.97 (2H, m, CH₂Me), 4.76 (1H, t, $J_{\text{CH}_2, \text{Me}}$ =7.0 Hz, 2-H), 6.98–7.03 (2H, m, Ar-H), 7.18–7.22 (2H, m, Ar-H).

4.2.4. 2,3-Dihydro-1,4-benzo[d]dithiine 4A. Yellow oil; $(R_f \ 0.4, \text{hexane})$; lit.²¹ 82.5–85 °C/0.18 mmHg; δ_{H} $(500 \text{ MHz}, \text{CDCl}_3)$ 3.26 (4H, s, 2 \times CH₂S), 6.97–7.00 (2H, dd, $J=5.9$, 3.4 Hz, 2 \times Ar-H), 7.13–7.17 (2H, dd, $J=5.9$, 3.4 Hz, $2\times$ Ar-H).

4.2.5. 3,4-Dihydro-2 H -1 λ ⁴,5-benzodithiepin 5A. White solid; mp 60–61 °C (from CHCl₃); lit.^{[22](#page-10-0)} 59–60 °C; $\delta_{\rm H}$ (500 MHz; CDCl₃) 2.30 (2H, m, CH₂CH₂CH₂), 2.86 (4H, m, $CH_2CH_2CH_2$), $7.15-7.18$ (2H, dd, $J=5.7$, 3.5 Hz, 2 \times Ar-H), $7.62 - 7.64$ (2H, dd, $J=5.6$, 3.5 Hz, 2 \times Ar-H).

4.2.6. 1,4-Benzo[d]dithiine 6A. Light pink coloured oil; bp 58–60 °C/0.03 mmHg; lit.^{[23](#page-10-0)} 67–70 °C/0.1 mmHg; δ_H (500 MHz, CDCl₃) 6.52 (2H, s, 2×CHS), 7.19-7.23 (2H, dd, $J=5.8$, 3.4 Hz, 2 \times Ar-H), 7.26–7.29 (2H, dd, $J=5.8$, 3.4 Hz, $2\times$ Ar-H).

4.2.7. 2,3-Dihydrobenzo[b]thiophene 7A. Colourless oil; bp 44–46 °C/0.15 mmHg; lit.^{[24](#page-10-0)} 67–69 °C/2 mmHg; $\delta_{\rm H}$ $(500 \text{ MHz}, \text{CDCl}_3)$ 3.23–3.27 (2H, m, SCH₂), 3.30–3.34 (2H, m, CH₂Ar), 6.90–6.93 (1H, ddd, $J_{5,4} = J_{5,6} = 7.4$ Hz, $J_{5,7}$ =1.0 Hz, 5-H), 7.00-7.04 (1H, ddd, $J_{6,5}$ = $J_{6,7}$ =7.4 Hz, $J_{6,4}$ =0.6 Hz, 6-H), 7.08–7.10 (1H, ddd, $J_{4,5}$ =7.4 Hz, $J_{4,6}$ = 0.6 Hz, 4-H), 7.12–7.13 (1H, dd, $J_{7.6}$ =7.4 Hz, $J_{7.5}$ =1.0 Hz, 7-H).

4.2.8. 1,3-Benzoxathiole 8A. Colourless liquid; bp 36° C/ 0.05 mmHg; lit.^{[25](#page-10-0)} 93-95 °C/5 mmHg; δ_{H} (500 MHz, CDCl3) 5.66 (2H, s, CH2), 6.81–6.83 (1H, ddd, $J_{7.6}$ =8.0 Hz, $J_{7.5}$ =1.2 Hz, $J_{7.4}$ =0.4 Hz, 7-H), 6.85–6.89 (1H, ddd, $J_{6,7}$ =8.0 Hz, $J_{6,5}$ =7.0 Hz, $J_{6,4}$ =1.2 Hz, 6-H), 6.97–7.00 (1H, ddd, $J_{5,4}$ =7.7 Hz, $J_{5,6}$ =7.6 Hz, $J_{5,7}$ = 1.2 Hz, 5-H), $7.15-7.17$ (1H, ddd, $J_{4,5}=7.7$ Hz, $J_{4,6}$ =1.2 Hz, $J_{4,7}$ =0.4 Hz, 4-H).

Sulfoxides $1B,^{10}$ $1B,^{10}$ $1B,^{10}$, $2B_{cis},^{10}$, $2B_{trans},^{10}$, $3B_{cis},^{10}$, $3B_{trans},^{10}$, $6B,^{11}$ $6B,^{11}$ $6B,^{11}$ $7B^{26,27}$ $7B^{26,27}$ $7B^{26,27}$ and $8B^{25}$ $8B^{25}$ $8B^{25}$ synthesised by the literature methods, were found to be spectrally identical to the metabolites isolated during the study.

4.3. Biotransformation of substrates 1A–8A using P. putida UV4, P. putida 8859 and P. putida 9816/11 to yield sulfoxides 1B–8B

Biotransformations of the bicyclic substrates 1A–8A were carried out with whole cell preparations of P. putida UV4 (TDO) and P. putida 8859 (NDO) and P. putida 9816/11 (NDO), using both small-scale $(<10 \text{ g})$ shake-flask and 10 l fermenter $(>10 \text{ g})$ conditions reported earlier for the sulfoxidation of both acyclic^{[5](#page-10-0)} and cyclic sulfides.^{[2](#page-10-0)} The workup procedure involved extraction of the culture medium using ethyl acetate.^{[2,5](#page-10-0)} Yields, chiroptical and ¹H NMR spectral data, obtained for each biotransformation, are summarised below.

4.3.1. 1,3-Benzodithiole 1A. (i) TDO product: (S)-1,3 benzodithiole-1-oxide 1B; white crystalline solid, 22 mg, 20% yield; $\alpha|_{D} = -170$ (c 1.0, CHCl₃); lit.^{[10](#page-10-0)} $\alpha|_{D} = -505$ (EtOH); 33% ee (CSPHPLC, Chiralcel OB, $\alpha=1.4$); $\delta_{\rm H}$ (500 MHz, CDCl₃) 4.18 (1H, d, $J_{A,B}$ =13.0 Hz, C H_A H_B), 4.33 (1H, d, $J_{B,A}$ =13.0 Hz, CH_AH_B), 7.30–7.33 (1H, m, Ar-H), 7.48-7.54 (2H, m, 2×Ar-H), 7.89-7.90 (1H, m, Ar-H).

(ii) NDO (8859) product: (R) -1,3-benzodithiole-1-oxide **1B**; 400 mg, 74% yield; $\alpha_{\text{D}} = +504$ (c 1.9, CHCl₃); $>98\%$ ee (CSPHPLC). A 10 litre fermenter (15 g substrate) gave sulfoxide 1B, 12 g, 72% yield; $[\alpha]_D = +498$ (c 1.9, CHCl₃) .98% ee (CSPHPLC).

(iii) NDO (9816/11) product: (R) -1,3-benzodithiole-1-oxide **1B**; 60 mg, 54% yield; $[\alpha]_D$ =+401 (c 1.3, CHCl₃); 81% ee (CSPHPLC).

4.3.2. 2-Methyl-1,3-benzodithiole 2A. (i) TDO products: $(1S, 2R)$ -2-methyl-1,3-benzodithiole-1-oxide $2B_{cis}$; white crystalline solid, 2.21 g, 40% yield; $[\alpha]_D = -319$ (c 0.7 CHCl₃); >98% ee (CSPHPLC, Chiralcel OD, $\alpha=1.33$); lit.^{[10](#page-10-0)} [α]_D=-68 (EtOH) for 25% ee; $\delta_{\rm H}$ (500 MHz, CDCl₃) 1.83 (3H, d, $J_{\text{Me,H}}$ =6.9 Hz, Me), 4.44 (1H, q, $J_{\text{H.Me}}$ =6.9 Hz, CHMe), 7.26–7.30 (1H, ddd, $J_{5,4}$ =7.8 Hz, $J_{5,6}$ =7.0 Hz, $J_{5,7}$ =1.0 Hz, 5-H), 7.43–7.46 (2H, m, 4-H and 6-H), 7.84– 7.86 (1H, dd, $J_{7.6}$ =7.2 Hz, $J_{7.5}$ =1.0 Hz, 7-H) and (1S,2S)-2methyl-1,3-benzodithiole-1-oxide $2B_{trans}$; 290 mg, 5% yield; $[\alpha]_D = -57$ (c 0.5, CHCl₃); $>98\%$ ee (CSPHPLC, Chiralcel OD, $\alpha=1.14$); lit.^{[10](#page-10-0)} $[\alpha]_D = -29$ (EtOH) for 25% ee; δ_H (500 MHz, CDCl₃) 1.60 (3H, d, $J_{Me,H}$ =7.4 Hz, Me), 4.59 (1H, q, $J_{H M e}$ =7.4 Hz, CHMe), 7.28–7.33 (1H, m, 5-H), 7.47–7.48 (2H, m, 4-H and 6-H), 7.83–7.85 (1H, dd, $J_{7,6}$ =7.7 Hz, $J_{7,5}$ =0.7 Hz, 7-H). When repeated on a 10 litre fermenter-scale (10.0 g of substrate $2A$), 1S,2R)-2-methyl-1,3- benzodithiole-1-oxide $2B_{cis}$; 2.9 g, 26% yield; $[\alpha]_D = -316$ (c 0.7 CHCl₃); >98% ee (CSPHPLC) and (1S,2S)-2-methyl-1,3-benzodithiole-1-oxide $2B_{trans}$; 66 mg, 1% yield; $[\alpha]_D = -60$ (c 0.7, CHCl₃); >98% ee (CSPHPLC) were obtained.

(ii) NDO (8859) products: $(1R,2S)$ -2-methyl-1,3-benzodithiole-1-oxide $2B_{cis}$; 2.21 g, 40% yield; $[\alpha]_D = +256$ (c 1.8, CHCl₃); 82% ee (CSPHPLC) and $(1R, 2R)$ -2-methyl-1,3-benzodithiole-1-oxide $2B_{trans}$; 390 mg, 7% yield; $[\alpha]_D = +21$ (c 0.7, CHCl₃); >38% ee (CSPHPLC).

4.3.3. 2-Ethyl-1,3-benzodithiole 3A. (i) TDO products: $(1S, 2R)$ -2-ethyl-1,3- benzodithiole-1-oxide $3B_{cis}$; white crystalline solid, 165 mg, 55% yield; $\lceil \alpha \rceil_D = -189$ (c 0.5, CHCl₃); >98% ee (CSPHPLC, Chiralcel OD, α =1.36); δ _H $(300 \text{ MHz}, \text{CDCl}_3)$ 1.30 (3H, t, $J_{\text{Me}, \text{CH}_2} = 7.4 \text{ Hz}, \text{ CH}_2 \text{Me}$), 2.05–2.16 (1H, m, $CH₂Me$), 2.33–2.44 (1H, m, $CH₂Me$), 4.26 (1H, dd, $J_{H,CH2}$ =7.6 Hz, 2-H), 7.25–7.28 (1H, m, Ar-H), $7.45 - 7.46$ (2H, m, Ar-H), $7.84 - 7.86$ (1H, d, $J=7.7$ Hz, Ar-H) and $(1R,2R)$ -2-ethyl-1,3-benzodithiole-1-oxide $3B_{trans}$; white crystalline solid, 15 mg, 5% yield; $[\alpha]_D = +24$ (c 0.4, CHCl₃); 18% ee (CSPHPLC, Chiralcel OD, $\alpha=1.11$); δ_H (300 MHz, CDCl₃) 1.18 (3H, t,

 $J_{\text{Me},\text{CH}_2}$ =7.4 Hz, CH₂Me), 1.65 (1H, m, CH₂Me), 2.00 (1H, m, C \tilde{H}_2 Me), 4.48 (1H, dd, $J_{2,1'A} = 8.7$ Hz, $J_{2,1'B} = 6.2$ Hz, H-2), 7.26–7.32 (1H, m, Ar-H), 7.47 (2H, m, Ar-H), 7.87 $(1H, d, J=7.7 Hz, Ar-H).$

4.3.4. 2,3-Dihydro-1,4-benzo[d]dithiine 4A. (i) TDO product: (R) -2,3-dihydrobenzo[d]dithiin-1-oxide 4B; white solid, 14 mg, 13% yield; mp $104-105$ °C (from CHCl₃); $[\alpha]_D = +37$ (c 1.3, CHCl₃); HRMS found: M⁺, 184.0016, $C_8H_8OS_2$ requires 184.0017; δ_H (500 MHz, CDCl₃) 3.05 (1H, ddd, $J_{3A,3B}$ =13.4 Hz, $J_{3A,2A}$ =6.6 Hz, $J_{3A,2B}$ =3.8 Hz, SCH_AH_B), 3.13 (1H, ddd, $J_{3B,3A}$ =13.4 Hz, $J_{3B,2A}$ =10.8 Hz, $J_{3B,2B}$ =3.6 Hz, SCH_AH_B), 3.40 (1H, ddd, $J_{2A,2B}$ =13.4 Hz, $J_{2A,3A}$ =6.6 Hz, $J_{2A,3B}$ =3.6 Hz, SOC H_A H_B), 3.76 (1H, ddd, $J_{2B,2A}$ =13.4 Hz, $J_{2B,3A}$ =10.8 Hz, $J_{2B,3B}$ =3.4 Hz, SOCH_AH_B), 7.25–7.28 (1H, ddd, $J_{6,5} = 8.2$ Hz, $J_{6,7} =$ 7.5 Hz, $J_{6.8}$ =1.2 Hz, 6-H), 7.31–7.32 (1H, dd, $J_{5.6}$ = 8.2 Hz, $J_{5,7}$ =1.2 Hz, 5-H), 7.37–7.40 (1H, ddd, $J_{7,8}$ = 7.8 Hz, $J_{7.6}$ =7.5 Hz, $J_{7.5}$ =1.2 Hz, 7-H), 7.73–7.75 (1H, dd, $J_{8,7}$ =7.8 Hz, $J_{8,6}$ =1.2 Hz, 8-H); >98% ee (CSPHPLC, Chiralcel OB, $\alpha=1.4$).

(ii) NDO (8859) product: (S) -2,3-dihydrobenzo[d]dithiin-1oxide 4B; 85 mg, 78% yield; α _D=-35 (c 1.1, CHCl₃); .98% ee (CSPHPLC).

(iii) NDO $(9816/11)$ product: (S) -2,3-dihydrobenzo[d]dithiin-1-oxide 4B; 67 mg, 62% yield; $[\alpha]_D = -37$ (c 1.3, CHCl₃); $>98\%$ ee (CSPHPLC); CD (λ , nm) 313 ($\Delta \epsilon$ -0.375), 271 ($\Delta \varepsilon$ 0.240), 256 ($\Delta \varepsilon$ -2.27), 242 ($\Delta \varepsilon$ 0.561), 229 ($\Delta \epsilon$ -2.815), 213 ($\Delta \epsilon$ -5.865), 194.2 ($\Delta \epsilon$ 2.100).

4.3.5. 3,4-Dihydro-2H-1 λ^4 ,5-benzodithiepin 5A. (i) NDO (8859) product: $(S)-1,2,3,4$ -tetrahydro-1 λ^4 ,5-benzodithiepin-1-oxide 5B; colourless crystalline solid, 30 mg, 26% yield; mp $120-121$ °C (from CHCl₃/hexane); $[\alpha]_D = -71$ (c 1.4, CHCl₃); Found: C, 54.7; H, 5.0; $C_9H_{10}OS_2$ requires C, 54.5; H, 5.1%; δ_H (500 MHz, CDCl3) 2.41–2.49 (2H, m, 4-H), 2.61–2.63 (1H, m, 3-H_A), 2.87–2.88 (1H, m, 3-H_B), 2.97–3.00 (1H, m, 2-H_A), 3.25–3.27 (1H, m, 2-H_B), 7.38–7.41 (1H, td, $J_{7,6}$ = $J_{7,8}$ =7.5 Hz, $J_{7,9}$ =1.4 Hz, 7-H), 7.58–7.59 (2H, m, 6-H and 8-H), $7.87 - 7.89$ (1H, dd, $J_{9,8} = 7.7$ Hz, $J_{9,7} = 1.4$ Hz, 9-H); 80% ee (CSPHPLC, Chiralcel OB, $\alpha=1.25$).

(ii) NDO (9816/11) product: $(S)-1,2,3,4$ -tetrahydro-1 λ^4 ,5benzodithiepin-1-oxide 5B; 20 mg, 10% yield; $\lceil \alpha \rceil_D = -82$ (c 0.8, CHCl₃); 93% ee (CSHPLC, Chiralcel OB, α =1.25); CD (λ , nm) 265 ($\Delta \varepsilon$ -2.162), 250 ($\Delta \varepsilon$ 0.439), 236 ($\Delta \varepsilon$ -1.243), 228 ($\Delta \varepsilon$ 0.448), 220 ($\Delta \varepsilon$ -1.423), 215 ($\Delta \varepsilon$ -0.589 , 198 ($\Delta \epsilon$ -7.96).

4.3.6. X-ray crystal structure analysis of (S)-1,2,3,4 tetrahydro-1 λ^4 ,5-benzodithiepin-1-oxide 5B. Recrystallization of compound $5B$ (93% ee) gave a sample whose X-ray crystal structure analysis showed it to be of $>98\%$ ee and of the (S) configuration.

Crystal data for 5B: $C_9H_{10}OS_2$, M_r =198.3, orthorhombic, $a=8.106(4)$, $b=9.043(3)$, $c=13.042(6)$ Å, $V=956.0(7)$ Å³, T=293 K, Cu K α radiation, λ =1.54178 Å, space group $P2_12_12_1$, $Z=4$, $D_x=1.378$ g cm⁻³, 0.50×0.40×0.34 mm³, μ =4.63 mm⁻¹, $F(000)$ =416, Siemens P3/V2000 diffractometer, ω scan, $10<2\theta<110^\circ$, measured/independent reflections: 5093/1195, direct methods solution, full matrix least squares refinement on F_0^2 , anisotropic displacement parameters for non-hydrogen atoms, hydrogens located in difference Fourier but included at positions calculated from the geometry of the molecule using the riding model, $R1=0.044$ for 1115 data with $F_0 > 4\sigma(F_0)$, 111 parameters, $wR2=0.113$ (all data), $GoF=1.07$, Flack absolute structure parameter $x=-0.09(4)$ establishes the absolute configuration as (1S), $\Delta\rho_{\text{min,max}}$ = -0.30/0.23 e Å⁻³. CCDC reference number 212386, Crystallographic data (excluding structure factors) for the structures in this paper have been deposited with the Cambridge Crystallographic Data Centre. Copies of the data can be obtained, free of charge, on application to CCDC, 12 Union Road, Cambridge, CB2 1EZ, UK [fax:+44(0)-1223-336033 or e-mail: deposit@ccdc.cam.ac.uk].

4.3.7. 1,4-Benzo[d]dithiine 6A. (i) TDO product: (S)-1,4 benzo[d]dithiin-1-oxide 6B; white solid, 25 mg, 23% yield; $[\alpha]_D = -376$ (c 0.5, CHCl₃); lit.^{[11](#page-10-0)} $[\alpha]_D = -383$ (CHCl₃); >98% ee (CSPHPLC, Chiralcel OB, $\alpha=1.17$); δ_{H} $(500 \text{ MHz}, \text{CDCl}_3)$ 7.05 (1H, d, $J_{3,2}$ =9.4 Hz, SCH), 7.31 (1H, d, $J_{2,3}$ =9.4 Hz, S(O)CH), 7.51–7.54 (1H, ddd, $J_{6,5}$ = $J_{6,7}$ =7.6 Hz, $J_{6,8}$ =1.4 Hz, 6-H), 7.56–7.60 (1H, ddd, $J_{7.6}$ = $J_{7.8}$ =7.6 Hz, $J_{7.5}$ =1.3 Hz, 7-H), 7.60–7.62 (1H, dd, $J_{5.6}$ =7.6 Hz, $J_{5.7}$ =1.3 Hz, 5-H), 7.94–7.96 (1H, dd, $J_{8.7}$ = 7.6 Hz, $J_{8.6}$ =1.4 Hz, 8-H); CD (λ , nm) 311 ($\Delta \epsilon$ 0.522), 272 $(\Delta \epsilon$ -0.444), 255 ($\Delta \epsilon$ 3.357), 243 ($\Delta \epsilon$ -0.675), 229 $(\Delta \epsilon + 3.657), 213 (\Delta \epsilon 7.470).$

(ii) NDO (8859) product: (R) -benzo[d]dithiin-1-oxide 6B; 66 mg, 60% yield; $[\alpha]_D = +373$ (c 1.0, CHCl₃); >98% ee (CSPHPLC).

(iii) NDO (9816/11) product: (R) -benzo[d]dithiin-1-oxide **6B**; 47 mg, 43% yield; $\alpha|_{D} = +373$ (c 1.0, CHCl₃); >98% ee (CSPHPLC).

4.3.8. 2,3-Dihydrobenzo[b]thiophene 7A. (i) TDO product: (R) -2,3-dihydrobenzo[b]thiophene-1-oxide 7B; white solid, 40 mg, 5% yield; $[\alpha]_D = -59$ (c 1.0, CHCl₃); lit.^{[27](#page-10-0)} $[\alpha]_D = -285$ (Me₂CO); 26% ee (CSPHPLC, Chiralcel OB, $\alpha=1.7$); $\delta_{\rm H}$ (500 MHz, CDCl₃) 3.23–3.26 (1H, m, 3-H_A), $3.28 - 3.34$ (1H, m, $3-H_B$), $3.39 - 3.40$ (1H, m, $2-H_B$), $3.83 -$ 3.91 (1H, m, 2-H_A), 7.41–7.44 (1H, ddd, $J_{5,4} = J_{5,6} = 7.4$ Hz, $J_{5,7}$ =0.5 Hz, 5-H), 7.46–7.47 (1H, dd, $J_{4,5}$ =7.4 Hz, $J_{4,6}$ = 1.2 Hz, 4-H), 7.50–7.53 (1H, ddd, $J_{6,5} = J_{6,7} = 7.4$ Hz, $J_{6,4} =$ 1.2 Hz, 6-H), 7.83–7.85 (1H, dd, $J_{7.6}$ =7.4 Hz, $J_{7.5}$ 0.5, 7-H).

(ii) NDO (8859) product: $(S)-2,3$ -dihydrobenzo[b]thiophene-1-oxide 7B; 48 mg, 21% yield; $[\alpha]_D = +95$ (c 1.1, CHCl₃); 38% ee (CSPHPLC); CD (λ, nm) 263 ($\Delta \epsilon$ 0.882), 231 ($\Delta \epsilon$ 0.733), 217 ($\Delta \epsilon$ -2.586), 198 ($\Delta \epsilon$ 5.447).

(iii) NDO (9816/11) product: (S) -2,3-dihydrobenzo $[b]$ thiophene-1-oxide 7B; 19 mg, 17% yield; $[\alpha]_D = +27$ (c 1.0, CHCl₃), 11% ee (CSPHPLC).

4.3.9. 1,3-Benzoxathiole 8A. (i) TDO product: (R) benzo[1,3]oxathiol-3-oxide **8B**; white solid, 11 mg, 11% yield; $[\alpha]_D = +194$ (c 1.0, CHCl₃); >98% ee (CSPHPLC, Chiralcel OB, $\alpha=1.3$); δ_H (500 MHz, CDCl₃) 4.98 (1H, d, $J_{A,B}$ =11.0 Hz, CH_AH_B), 5.49 (1H, d, $J_{B,A}$ =11.0 Hz, CH_AH_B), 7.18–7.20 (1H, ddd, $J_{4,5}$ =7.6 Hz, $J_{4,6}$ =1.2 Hz, $J_{4,7}$ =0.5 Hz, 4-H), 7.22–7.24 (1H, ddd, $J_{5,4}$ =7.6 Hz, $J_{5,6}$ =7.2 Hz, $J_{5,7}$ =1.2 Hz, 5-H), 7.56–7.59 (1H, ddd, $J_{6,7}$ =7.7 Hz, $J_{6,5}$ =7.2 Hz, $J_{6,4}$ =1.2 Hz, 6-H), 7.88–7.90 (1H, ddd, $J_{7.6}$ =7.7 Hz, $J_{7.5}$ =1.2 Hz, $J_{7.4}$ =0.5 Hz, 7-H); CD (λ, nm) 285 ($\Delta \epsilon$ -2.831), 215 ($\Delta \epsilon$ 6.55), 208 ($\Delta \epsilon$ 5.259), 199 ($\Delta \varepsilon$ 15.600).

(ii) NDO (8859) product: (R) -benzo $[1,3]$ oxathiol-3-oxide **8B**; 50 mg, 2% yield; $[\alpha]_D = +87$ (c 0.5, CHCl₃); 43% ee (CSPHPLC)

(iii) NDO $(9816/11)$ product: (R/S) -benzo $[1,3]$ oxathiol-3oxide 8B; 200 mg, 70% yield; $[\alpha]_D = 0$ (c 0.5, CHCl₃); racemic (CSPHPLC).

4.4. Reaction of $(+)$ - (R) -benzo-1,3-dithiole-1-oxide 1B with n-butyllithium and benzaldehyde

n-Butyllithium solution in hexane (1.6 M, 1.6 ml) was added to a stirring solution of $(+)$ - (R) -benzo-1,3-dithiole-1oxide 1B (300 mg, 1.7 mmol in 10 ml of dry THF) kept under nitrogen at -78 °C. After 0.5 h, benzaldehyde (186 mg, 1.7 mmol) was added to the reaction mixture; it was allowed to warm to room temperature and stirred for another 0.5 h. The reaction mixture was then carefully poured into water (50 ml), the products extracted with ether $(3\times25 \text{ ml})$, the extract washed with water (30 ml) and dried (MgSO4). Removal of the solvent yielded the crude product mixture which on separation by PLC $(1\% \text{ MeOH in CHCl}_3)$ afforded pure samples of $cis-(1R,2S,1/S)-2-(1'-phenyl$ methanol)-benzo-1,3-dithiole-1-oxide 10 and the more polar trans-(1R,2R,1'R)-2-(1'-phenylmethanol)-benzo-1,3dithiole-1-oxide 9.

4.4.1. $(1R, 2S, 1'S)$ -2- $(1'$ -Phenylmethanol)-benzo-1,3dithiole-1-oxide 10. Colourless crystals, 147 mg, 31% yield; R_f 0.31 (1% MeOH in CHCl₃); mp 163-164 °C (from CHCl₃/hexane); $[\alpha]_D = +344$ (c 0.49, CHCl₃); HRMS found: M⁺276.0278, C₁₄H₁₂O₂S₂ requires 276.0278; $\delta_{\rm H}$ (300 MHz, CDCl₃) 4.23 (1H, d, $J_{OH,1'}$ 1.7, OH), 4.65 (1H, d, $J_{2,1}$ =4.8 Hz, 2-H), 5.77 (1H, d, $J_{2,1}$ =4.8 Hz, 1[']-H), 7.25– 7.28 (1H, m, Ar-H), 7.36–7.46 (5H, m, Ar-H), 7.54 (2H, d, $J=7.1$ Hz, Ar-H), 7.77 (1H, d, $J=7.6$ Hz, Ar-H); m/z 276 $(M⁺, 7%)$, 259 (13), 229 (30) and 153 (100).

4.4.2. X-ray crystal structure analysis of $(1R, 2S, 1'S)$ -2-(1'-phenylmethanol)-benzo-1,3-dithiole-1-oxide 10. Crystal data for **10**: $C_{14}H_{12}O_2S_2$, $M_r=276.4$, orthorhombic, $a=5.531(1), b=10.410(3), c=21.931(7)$ Å, $V=1262.8(6)$ Å³, T=293 K, Cu K α radiation, λ =1.54178 Å, space group $P2_12_12_1$, $Z=4$, $D_x=1.45$ g cm⁻³, 0.56×0.49×0.43 mm³, μ =3.74 mm⁻¹, Siemens P3/V2000 diffractometer, ω scan, $8 < 2\theta < 150^{\circ}$, measured/independent reflections: 2748/ 2303, direct methods solution, full matrix least squares refinement on F_0^2 , anisotropic displacement parameters for non-hydrogen atoms, hydrogens located in difference Fourier but included at positions calculated from the geometry of the molecule using the riding model, $R1=0.071$ for 2181 data with $F_0 > 4\sigma(F_0)$, 165 parameters, wR2=0.188 (all data), $GoF=1.05$, Flack absolute structure parameter $x=0.00(4)$ establishes the absolute configuration as $(1R, 2S, 1'S)$,

 $\Delta \rho_{\text{min,max}}$ = -0.47/0.54e \AA ⁻³. CCDC reference number 212387.

4.4.3. $(1R, 2R, 1'R)$ -2- $(1'$ -Phenylmethanol)-benzo-1,3dithiole-1-oxide 9. White solid, 108 mg, 23% yield; R_f 0.19 (1% MeOH in CHCl₃); $[\alpha]_D = +116$ (c 1.2, CHCl₃); HRMS found: $M^+276.0291$, $C_{14}H_{12}O_2S_2$ requires 276.0278; $\delta_{\rm H}$ (300 MHz; CDCl₃) 4.68 (1H, d, $J_{2,1}$) 10.0 Hz, 2-H), 4.93 (1H, br s, OH), 5.32 (1H, d, $J_{1',2}$ =10.0 Hz, 1'-H), 7.22–7.44 (6H, m, Ar-H), 7.49–7.52 $(2H, m, Ar-H), 7.83$ (1H, d, J=7.8 Hz, Ar-H); m/z 276 (M⁺, 4%), 259 (11), 229 (28) and 153 (100).

4.4.4. Deoxygenation of $(1R, 2S, 1'S)$ -2- $(1'$ -phenylmethanol)-benzo-1,3-dithiole-1-oxide (10) and $(1R, 2R, 1/R)$ -2-(1'-phenylmethanol)-benzo-1,3-dithiole-1-oxide 9. To each sample of monosulfoxides 10 and 9 (20 mg, 0.72 mmol), dissolved separately in CH_2Cl_2 (3 ml), was added triphenyl phosphine (25 mg, 0.1 mmol) and a catalytic amount of trichlorooxobis(triphenyl phosphine) rhenium. The two reaction mixtures were allowed to stir at room temperature (sulfoxide 9 for 2 h and sulfoxide 10 for 72 h). The products were purified by PLC to give the corresponding sulfides $(+)$ - $(1'S)$ -11; 8 mg, 44% yield; $[\alpha]_D = +12$ (c 0.75, Me₂CO) from 1R,2S,1⁷S-10 and (-)- $(1\ddot{R})$ -11; 9 mg, 49% yield; $[\alpha]_D$ =-12 (c 0.9, Me₂CO) from $1R, 2R, 1'R - 9$); HRMS found: M⁺260.0342, C₁₄H₁₂OS₂ requires 260.0330; $\delta_{\rm H}$ (300 MHz, CDCl₃) 4.66 (1H, d, $J=8.2$ Hz, 2-H), 4.89 (1H, d, $J=8.2$ Hz, $1'$ -H), 7.03-7.06 (2H, m, Ar-H), 7.18–7.25 (3H, m, Ar-H), 7.32–7.40 (5H, m, Ar-H); m/z 260 (M⁺, 9%) and 153 (100).

4.4.5. Sodium periodate oxidation of $(+)$ - (R) -1.3-benzodithiole-1-oxide 1B. $(+)$ - (R) -1,3-Benzodithiole-1-oxide 1B (260 mg, 1.52 mmol) in methanol (15 ml) was oxidized using a solution of sodium periodate (360 mg, 1.68 mmol) in water (10 ml) at 50 °C for 24 h. PLC (5% MeOH in $CHCl₃$), of the reaction mixture, afforded pure samples of $(1R,3R)-1,3$ -benzodithiole-1,3-dioxide $1C_{trans}$ and the more polar cis-1,3-benzodithiole-1-oxide $1C_{cis}$.

 $(1R,3R)$ -1,3-Benzodithiole-1,3-dioxide $1C_{trans}$. White crystalline solid, 156 mg, 75% yield; R_f 0.33 (5% MeOH in CHCl₃); mp $123-126$ °C (from CHCl₃/hexane); $[\alpha]_D = +646$ (c 1.5, CHCl₃); HRMS found: M⁺185.9810, $C_7H_6O_2S_2$ requires 185.9809; δ_H (300 MHz; CDCl₃) 4.46 (2H, s, CH2), 7.82–7.85 (2H, m, Ar-H), 7.99–8.02 (2H, m, Ar-H); m/z 186 (M⁺, 32%) and 156 (100). Benzo-1,3dithiole-1,3-dioxide $1C_{cis}$. 40 mg, 19% yield; R_f 0.22 (5%) MeOH in CHCl₃); mp 179-180 °C; HRMS found: M^+ 185.9814, C₇H₆O₂S₂ requires 185.9809; δ_H (300 MHz, CDCl₃) 4.24 (1H, d, $J_{A,B} = 13.2$ Hz, CH_AH_B), 5.03 (1H, d, $J_{\text{B,A}}$ =13.2 Hz, CH_AH_B), 7.85–7.88 (2H, m, Ar-H), 8.08– 8.11 (2H, m, Ar-H); m/z 186 (M⁺, 33%) and 156 (100).

4.5. Ring opening reactions of bicyclic disulfoxide $1C_{trans}$ and monosulfoxides 7B and 8B with alkyl and aryllithium reagents

4.5.1. Reaction of (1R,3R)-1,3-benzodithiole-1,3-dioxide $1C_{trans}$ with *n*-butyllithium and benzaldehyde. *n*-Butyllithium solution in hexane (1.6 M, 0.53 ml) was added to stirred solution (1R,3R)-1,3-benzodithiole-1,3-dioxide

1C_{trans} (100 mg, 0.54 mmol, $[\alpha]_D$ =+646) in dry THF at -78 °C. After 0.5 h, benzaldehyde (0.06 ml, 0.6 mmol) was added to the reaction mixture; it was allowed to warm to room temperature, stirred overnight, and worked up as described earlier. Purification by PLC (5% MeOH in CHCl₃) afforded a pure sample of $(-)$ -(butane-1-sulfinylmethanesulfinyl)-benzene 13; white solid, 64 mg, 49% yield; R_f 0.43 (5% MeOH in CHCl₃); mp 82–84 °C (from CH₂Cl₂/hexane); $[\alpha]_D = -180$ (c 1.69, CHCl₃); HRMS found: M⁺244.0582, C₁₁H₁₆O₂S₂ requires 244.0592; $\delta_{\rm H}$ $(500 \text{ MHz}, \text{ CDCl}_3)$ 0.93 (3H, t, J=7.5 Hz, $(\text{CH}_2)_3Me$), 1.46–1.53 (2H, m, $(CH_2)_2CH_2CH_3$), 1.75–1.82 (2H, m, CH₂CH₂CH₃), 3.03–3.09 (1H, m, CH₄H_R- $CH_2CH_2CH_2CH_3$), $(CH_2)_2CH_3$, 3.18–3.24 (1H, m, $CH_AH_B(CH_2)_2CH_3$), 3.91 (1H, d, J=13.3 Hz, SO–C $H_A H_B$ –SO), 4.15 (1H, d, $J=13.3$ Hz, SO–CH_AH_B–SO), 7.55–7.59 (3H, m, Ar-H), 7.68–7.71 (2H, m, Ar-H); m/z 244 (M⁺, 2%) and 126 (100); CD (λ , nm) 251 ($\Delta \epsilon$ -11.18), 220 ($\Delta \epsilon$ 16.67).

4.5.2. Reaction of $(+)$ - $(1R,3R)$ -benzo-1,3-dithiole-1,3dioxide with phenyllithium. General procedure. The lithium reagent (1.1 molar equiv.) was added to a stirring solution $(-78 \degree C,$ under nitrogen) of the disulfoxide $(1C_{trans})$ or mono-sulfoxide (7B, 8B) (1.0 mmol in) \sim 20 ml of dry THF). The reaction mixture was allowed to warm up to room temperature, stirred overnight, and then quenched with water; it was extracted with ethyl acetate, the extract dried $(MgSO₄)$, concentrated under reduced pressure, and the crude product purified by PLC.

 $(1R,3R)$ -Benzo-1,3-dithiole-1,3-dioxide $1C_{trans}$. (50 mg, 0.27 mmol, $[\alpha]_D = +646$ and phenyllithium solution in cyclohexane (1.8 M, 0.25 ml) yielded meso bis-(phenylsulfinyl) methane 14; white solid, 23 mg, 32% yield; mp 124–125 °C (from CHCl₃); lit.¹⁴ 123 °C; R_f 0.25 (CHCl₃); $\delta_{\rm H}$ (500 MHz; CDCl₃) 4.10 (1H, d, $J_{\rm A,B}$ =12.7 Hz,-SO– CH_AH_B-SO-), 4.20 (1H, d, $J_{B,A}=12.7$ Hz, –SO–CH_AH_B– SO–), 7.51–7.59 (6H, m, Ar-H), 7.68–7.74 (4H, m, Ar-H).

4.5.3. Reaction of $(+)$ - $(1R,3R)$ -benzo-1,3-dithiole-1,3dioxide $1C_{trans}$ with methyllithium. $(1R,3R)$ -Benzo-1,3dithiole-1,3-dioxide $1C_{trans}$ (45 mg, 0.24 mmol, $[\alpha]_{D}$ = $+646$) and methyllithium solution in hexane (1.6 M, 0.25 ml) yielded $(-)$ -(1R)-[(3S)-methylsulfinyl] methylphenyl sulfoxide 15; white solid, 38 mg, 59% yield; mp 135 $^{\circ}$ C (from CHCl₃); R_f 0.22 (CHCl₃); $[\alpha]_D = -234$ (c 0.8, CHCl₃); HRMS found: M⁺202.0118, C₈H₁₀O₂S₂ requires 202.0122; δ_H (500 MHz, CDCl₃) 2.98 (3H, s, Me), 3.91 (1H, d, $J_{A,B}$ =13.1 Hz, CH_AH_B), 4.13 (1H, d, $J_{B,A}$ =13.1 Hz, CH_AH_B), 7.55–7.60 (3H, m, Ar-H), 7.67–7.70 (2H, m, Ar-H); m/z: 202 (M⁺1%),139 (26), 125 (53), 109 (72), 91 (100); CD (λ , nm) 251 ($\Delta \epsilon$ -18.15), 220 ($\Delta \epsilon$ 20.42).

4.5.4. Deoxygenation of disulfoxide 15. To a solution of $(-)-(1R)$ - $(3S)$ -methylsulfinyl] methylphenyl sulfoxide 15 (22 mg, 0.11 mmol, $[\alpha]_D = -234$) in CH₂Cl₂ (3 ml) was added triphenylphosphine (25 mg, 0.1 mmol) along with a catalytic amount of trichlorooxobis(triphenylphosphine) rhenium; the reaction mixture was stirred (2 h) at room temperature. Triphenylphosphine oxide was removed from the reaction mixture by reverse phase PLC (70% MeOH in $H₂O$). The product was further separated into two sulfoxides by PLC on silca gel (5% MeOH in CHCl₃); $(-)$ - (S) -(phenylsulfanylmethyl)methyl sulfoxide 20; 8 mg, 40% yield; $\lceil \alpha \rceil_D = -27$ (c 0.81, CHCl₃); 24% ee (Chiralcel OJ) and $(-)$ - (R) -(methylsulfanylmethyl)phenyl sulfoxide 21; 5 mg, 22% yield; $[\alpha]_D = -110$ (c 0.45, CHCl₃); 87% ee (Chiralcel OD). Spectral data for sulfoxides 20 and 21 were in agreement with the literature values.⁶

4.5.5. Reaction of $(+)$ - $(1R,3R)$ -benzo-1,3-dithiole-1,3dioxide 1C_{trans} with *n*-hexyllithium. $(1R,3R)$ -Benzo-1,3-
dithiole-1,3-dioxide 1C_{trans} (40 mg, 0.22 mmol, dithiole-1,3-dioxide $1C_{trans}$ (40 mg, 0.22 mmol, $[\alpha]_D = +646$ and *n*-hexyllithium solution in hexane $(2.0 \text{ M}, 0.15 \text{ ml})$ yielded $(-)-(1R)-(3S)-n$ -hexylsulfinyl methylphenyl sulfoxide 16; white solid, 38 mg, 65% yield; mp $128-130$ °C (from benzene); R_f 0.2 (CHCl₃); $[\alpha]_D = -247$ (c 1.7, CHCl₃); HRMS found: M⁺272.0902, $C_{13}H_{20}O_{2}S_{2}$ requires 272.0905; δ_{H} (500 MHz, CDCl₃) 0.89 $(3H, t, J_{Me, CH_2} = 7.1 \text{ Hz}, \text{Me}), 1.30-1.34 \text{ (4H, m, (CH}_2)_{3}$ $(CH_2)_2$ Me), 1.45–1.47 (2H, m, $(CH_2)_2CH_2(CH_2)_2$ Me), 1.77–1.82 (2H, m, $CH_2CH_2(CH_2)$ ₃Me), 3.04–3.08 (1H, m, $CH_AH_B-(CH_2)_4Me$), 3.16–3.21 (1H, m, $CH_AH_B (CH_2)_4$ Me), 3.90 (1H, d, $J_{A'B'}=13.2$ Hz, SO–C $H_{A'}H_{B'}$ SO), 4.13 (1H, d, $J_{B'A'}=13.2$ Hz, SO-CH_A $H_{B'}$ -SO), 7.55–7.60 (3H, m, Ar-H), 7.68–7.71 (2H, m, Ar-H); m/z 272 (M^+ , 1%), 210 (5), 126 (81) and 43 (100); CD (λ , nm) 256 ($\Delta \epsilon$ -21.39), 221 ($\Delta \epsilon$ 23.55).

4.5.6. Reaction of $(+)$ - $(1R,3R)$ -benzo-1,3-dithiole-1,3dioxide $1C_{trans}$ with t-butyllithium. $(1R,3R)$ -Benzo-1,3dithiole-1,3-dioxide $1C_{trans}$ (60 mg, 0.32 mmol) and t-butyllithium solution in pentane $(1.7 M, 0.3 ml)$ yielded $(+)$ - $(1R)$ - $[(3S)$ -t-butylsulfinyl]methylphenyl sulfoxide 17 as a light yellow coloured oil, 20 mg, 30% yield; R_f 0.2 (CHCl₃); $[\alpha]_D = +75$ (c 1.0, CHCl₃); HRMS found: M⁺+H 245.0666, $C_{11}H_{16}S_2O_2+H$ requires 245.0670; δ_H (500 MHz; CDCl₃) 1.21 (9H, s, Bu^t), 3.91 (1H, d, J_{AB} =12.4 Hz, CH_AH_B), 3.95 (1H, d, J_{BA} =12.4 Hz, CH_AH_B), 7.58 (3H, m, Ar-H), 7.79 (2H, m, Ar-H); m/z (CI) 245 (M⁺+H, 7%), 189 (15), 126 (96) and 57 (100); CD (λ , nm) 250 ($\Delta \epsilon$ -14.66), 218 ($\Delta \epsilon$ 26.49).

4.5.7. Reaction of $(-)$ - (R) -2,3-dihydrobenzothiophene-1oxide $7B$ with *n*-butyllithium. *n*-Butyllithium solution in hexane $(1.6 \text{ M}, \quad 0.53 \text{ ml})$ and $(-)-(R)-2,3-\text{dihydro}$ benzo $[b]$ thiophene-1-oxide **7B** (90% ee) (118 mg, 0.78 mmol) yielded $(-)$ - (R) -n-butylphenethyl sulfoxide 18 as a colourless oil, 45 mg, 28% yield; R_f 0.51 (5% MeOH in CHCl₃); $[\alpha]_D = -13.3$ (c 1.98, CHCl₃); HRMS found: M⁺210.1072, C₁₂H₁₈OS requires 210.1078; $\delta_{\rm H}$ (500 MHz; CDCl₃) 0.95 (3H, t, $J_{1,2}$ =7.4 Hz, Me), 1.39–1.55 (2H, m, CH₂Me), 1.71–1.78 (2H, m, CH_EH_FCH₂Me), 2.60–2.66 (1H, ddd, $J_{C,D}$ =13.0 Hz, $J_{C,A}$ =9.0 Hz, $J_{C,B}$ =8.5 Hz, PhCH₂-H_CH_D), 2.70-2.76 (1H, ddd, J_{D,C}=13.0 Hz, $J_{\text{D,A}}$ =7.8 Hz, $J_{\text{D,B}}$ =5.9 Hz, PhCH₂-H_CH_D), 2.86–2.93 (1H, ddd, $J_{E,F}$ =12.8 Hz, $J_{E,G}$ =10.0 Hz, $J_{E,H}$ =7.2 Hz, SOC H_E H_F), 2.93–2.98 (1H, ddd, J_{F,E}=12.8 Hz, J_{F,G}= 9.3 Hz, $J_{\text{FH}}=5.6$ Hz, SOCH_FH_F), 3.04–3.10 (1H, ddd, $J_{A,B}$ =14.0 Hz, $J_{A,C}$ =9.0 Hz, $J_{A,D}$ =7.8 Hz, Ph–C H_A H_B), 3.11–3.17 (1H, ddd, $J_{B,A}$ =14.0 Hz, $J_{B,C}$ =8.5 Hz, $J_{\text{B,D}}$ =5.9 Hz, Ph–CH_AH_B), 7.23–7.26 (3H, m, Ar-H), 7.31–7.34 (2H, m, Ar-H); m/z (EI) 210 (M⁺, 13%), 194 (24), 193 (32), 154 (7), 135 (8), 105 (100), 91 (69), 77 (57), 57 (92), 41 (47), 29 (55).

4.5.8. Reaction of $(-)$ - (S) -1,3-benzoxathiole 8B with *n*-butyllithium. *n*-Butyllithium solution in hexane $(1.6 M,$ 0.16 ml) and $(-)$ - (S) -1,3-benzoxathiole-1-oxide 8B (73%) ee) (35 mg, 0.23 mmol) yielded $(+)$ - (R) - n -butylphenoxymethyl sulfoxide 19 as an oil; 16 mg, 34% yield; R_f 0.52 (5% MeOH in CHCl₃); $[\alpha]_D = +74$ (c 0.6, CHCl₃); HRMS found: M⁺, 212.0866, C₁₁H₁₆O₂S requires 212.0871; m/z 212 (Mþ, 70%), 108 (97), 107 (88), 94 (96), 79 (95), 77 (87), 65 (92), 57 (66), 55 (99), 51 (94), 41 (97), 39 (95), 29 (100), 27 (95); $\delta_{\rm H}$ (500 MHz; CDCl₃) 0.98 (3H, t, $J_{\rm CH_2,Me}$ =7.3 Hz, Me), $1.44-1.59$ (2H, m, $CH₂Me$), $1.78-1.85$ (2H, m, CH_2CH_2Me), 2.80–2.91 (2H, m, $CH_2CH_2CH_2Me$), 4.92 (1H, d, $J_{A,B}$ =10.2 Hz, OCH_AH_B), 5.02 (1H, d, $J_{B,A}$ = 10.2 Hz, OCH_AH_B), 7.02–7.08 (3H, m, Ar-H), 7.31–7.33 (2H, m, Ar-H); 49% ee (CSP-HPLC, Chiralcel OD Column $\alpha = 1.1$).

Acknowledgements

We thank the BBSRC, DTI, Avecia Lifesciences Molecules, Astra Zeneca and Oxford Asymmetry for financial support under the LINK Scheme (N. D. S.), DENI (S. H., S. S.) and the Queen's University of Belfast (M. K.) for postgraduate studentship.

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